

Waterbodies Eligible for Discharge Under Table B of the GPPC Renewal Permit*

Any existing or proposed discharges to the waterbodies specified below, or to waterbodies upstream of the waterbodies specified below, are ineligible for discharge under Table A, but may be eligible for discharge under Table B.

Note: Counties listed indicate the primary locations of the listed waterbodies; however, these waterbodies may flow from or into other bordering counties. Alternate or local spellings are included in “()”. Any discharges to portions of these waterbodies which are classified as FW1, C1 or PL waters are ineligible for authorization under Tables A or B.

Atlantic County

Absecon Creek and its unnamed tributaries
Jarretts Run
Doughty Pond
North Branch (Absecon Creek)
South Branch (Absecon Creek)

Mercer County

Millstone River and its unnamed tribs.
Delaware-Raritan Canal

Bergen County

Spark Hill (Sparkill) Creek
Hirshfeld Brook and its unnamed tributaries
Oradell Reservoir
Saddle River and its unnamed tributaries
Sprout Brook and its unnamed tributaries
Pehle Brook
Jordan Brook
Hohokus Brook and its unnamed tributaries
Allendale Brook
Ramsey Brook
Valentine Brook
Saddle Brook
Pine Brook
Haledon Reservoir
Long Swamp Brook
Burd Reservoir

Middlesex County

Millstone River and its unnamed tributaries
Heathcote Brook and its unnamed tributaries
Carters Brook and its unnamed tributaries
Heathcote Brook Branch
Harrys Brook
Lawrence Brook and its unnamed tribs.
Sawmill Brook and its unnamed tribs.
Sucker Brook
Beaver Dam Brook
Ireland Brook
Oakeys Brook and its unnamed tribs.
Cow Yard Brook
Terhune Run
Tennents Pond
Tennents Brook (Creek) and its unnamed tribs.
Warnes Brook
Matchaponix Brook and its unnamed tribs.
Barclay Brook and its unnamed tributaries

Essex County

Canoe Brook
Canoe Brook Reservoir
Taylor Brook
Bear Brook
Cub Brook and its unnamed tributaries
Orange Reservoir
East Branch Rahway River
West Branch Rahway River and its tribs.
Turtle Brook

Hunterdon County

Swan Creek Reservoir East
Swan Creek Reservoir West
Delaware-Raritan Canal

Waterbodies Eligible for Discharge Under Table B of the GPPC Renewal Permit* (contd.)

Monmouth County

Swimming River and its unnamed tribs.
Swimming River Reservoir
Pine Brook and its unnamed tributaries
Hop Brook and its unnamed tributaries
Willow Brook and its unnamed tributaries
Big Brook and its unnamed tributaries
Trout Brook
Yellow Brook and its unnamed tributaries
Slope Brook
Manasquan River and its unnamed tributaries
Manasquan Reservoir
Cooks Creek
Deep Creek
Crabtown Creek
Debbies Creek
Watson Creek
Judas Creek
Roberts Swamp Brook
Bayhead-Manasquan Canal
Sawmill Creek
Squankum Brook and its unnamed tribs.
Mingamahone Brook and its tribs.
Long Swamp Brook
E. Branch Mingamahone Brook and its tribs.
Bear Swamp Brook
Marsh Bog Brook and its unnamed tribs.
Bannen Meadow Brook
Cricket Creek
Cattail Brook
DeBois Creek and its unnamed tribs.
Burkes Creek
Applegates Creek
Shark River and its unnamed tribs.
Musquash Brook
Laurel Gully
Jumping Brook
Wills Brook
Hankins Brook
Robins Swamp Brook
Reevy Branch of Shark River
Matchaponix Brook and its unnamed tribs.
Pine Brook and its unnamed tribs.
Rocky Brook and its unnamed tribs.
Weamaconk Creek and its unnamed tribs.
McGellairds Brook and its unnamed tribs.

Morris County

Taylor Town Reservoir
Mine Brook and its tributaries
Lower Mine Hill Reservoir
Clyde Potts Reservoir
Beaver Creek (Brook)
Split Rock Reservoir
Burd Reservoir
Charlottesville Reservoir
Lake Kinnelon
Takeout Reservoir
Butler Reservoir

Ocean County

Metdeconk River and its unnamed tribs.
Cedar Bridge Brook and tribs.
South Branch (Metdeconk River)
North Branch (Metdeconk River)
Cottler Brook and its unnamed tribs.
Cabinfield/Schoolhouse Brook and its tribs.
Gravelly Run
Muddy Ford Brook and its tributaries
Snipe Creek and its tributaries
Turtle Brook and its tributaries
Ridge Creek
Watering Place Brook and its tributaries

Passaic County

Pequannock River and its unnamed tribs.
Oak Ridge Reservoir
Wanaque River and its unnamed tribs.
Wanaque Reservoir
Stone House Brook
Macopin River
Timber Brook
Pacock (Pacack) Brook and its unnamed tribs.
Charlottesville Reservoir
Point View Reservoir

Salem County

Salem Canal
Laurel Lake
Elkinton Mill Pond

**Waterbodies Eligible for Discharge Under Table B of the GPPC Renewal Permit*
(continued)**

Somerset County

Delaware-Raritan Canal
Millstone River and its tributaries
Royce Brook and its unnamed tributaries
Six Mile Run and its unnamed tributaries
Middlebush Brook and its unnamed tributaries
Ten Mile Run and its unnamed tributaries
Simonson Brook and its unnamed tributaries
Bedens Brook and its unnamed tributaries
Pike Run and its unnamed tributaries
Rock Brook and its unnamed tributaries

Sussex County

Dry Brook Reservoir
Franklin Pond
Franklin Pond Creek and its tributaries
Morris Lake (Newton Reservoir)
Lake Rutherford

Union County

Rahway River and its unnamed tributaries
upstream of Valley Rd.
Nomaheagan Brook and its unnamed tribs.
Turtle Brook

Counties of Union, Middlesex, Monmouth, Ocean, Burlington, Atlantic, Cape May, Cumberland
and Salem: Shellfish Harvesting Waters

The Department reserves the right to deny an individual authorization under the GPPC permit renewal to any proposed discharges to waterbodies identified in the Department's annual Shellfish Growing Classification Charts under the following shellfish growing water classification codes: special restricted areas; seasonal areas; prohibited areas and; approved areas. The designated uses for waterbodies classified as SE1 or SC include shellfish harvesting in accordance with N.J.A.C. 7:9B-1.1 *et seq.* If the Department determines that it can not approve a discharge to a waterbody used for shellfish harvesting under the GPPC permit renewal, the applicant will be notified of the Department's finding. The Department will also notify the applicant if the discharge can be authorized under an individual NJPDES DSW permit. If the Department can authorize a discharge to a waterbody classified as SE1 or SC under the GPPC permit renewal, the Department would issue such authorization under Table B.

All Counties

In issuing requests for authorization under the GPPC permit renewal, the Department may deem discharges to other waterbodies ineligible under Table A on a case-by-case basis. These circumstances may apply to a discharge proposed to a waterbody where this waterbody recharges potable groundwater wells downgradient of the proposed discharge. Or, information may become available to the Department which shows that a waterbody not included in this list is used for potable water purposes. If the Department determines that such site-specific circumstances apply and the conditions of Table A are not appropriate, the Department will notify the applicant prior to issuing an individual GPPC authorization under Table B.

Other Pollutants that may be Limited Under Table D

In addition to complying with the effluent limitations and monitoring conditions of Table D on the preceding pages, any other parameters indicated below may also be limited in an individual authorization. These additional parameters will be included in Part III of the individual authorization. All units are in µg/L.

| Parameter | FW2 Waters | | SE, SC Waters | | RQL* |
|-------------------------------|------------|-----------|---------------|-----------|------|
| | Monthly / | Daily | Monthly / | Daily | |
| | Avg. | / Maximum | Avg. | / Maximum | |
| Volatile Compounds | | | | | |
| Acrolein | NL | 100 | NL | 100 | 50 |
| Acrylonitrile | NL | 100 | NL | 100 | 50 |
| Bromoform | NL | 8.6 | 29 | 58 | 8 |
| Carbon Tetrachloride | NL | 6 | 8.8 | NL | 6 |
| Chlorobenzene | 15 | 28 | 15 | 28 | 6 |
| Chlorodibromomethane | NL | 8.2 | NL | 14 | 6 |
| Chlorethane | 104 | 268 | 104 | 268 | - |
| Chloroform | NL | 11.4 | 21 | 46 | 5 |
| Dichlorobromomethane | NL | 5 | NL | 12 | 5 |
| 1,1-Dichloroethane | 22 | 59 | 22 | 59 | - |
| 1,2-Dichloroethane | NL | 3 | 68 | 211 | 3 |
| 1, 1-Dichloroethylene | NL | 6 | 16 | 25 | 6 |
| 1,2-Dichloropropylene | 153 | 230 | 153 | 230 | - |
| 1,3-Dichloropropane | 10 | 20 | 29 | 44 | - |
| Methyl Bromide | 20 | 40 | 20 | 40 | 9 |
| Methyl Chloride | 86 | 190 | 86 | 190 | 10 |
| Methylene Chloride | NL | 9.4 | 40 | 89 | 6 |
| 1,1,2,2 Tetrachloroethane | NL | 10 | NL | 10 | 10 |
| Tetrachloroethylene | NL | 16 | 22 | 56 | 9 |
| 1,2-Trans-Dichloroethylene | 21 | 54 | 21 | 54 | 4 |
| 1,1,1-Trichloroethane | 21 | 54 | 21 | 54 | 6 |
| 1,1,2-Trichloroethane | NL | 12 | 21 | 54 | 6 |
| Trichloroethylene | NL | 5.4 | 21 | 54 | 5 |
| Vinyl Chloride | NL | 10 | 104 | 268 | 10 |
| Acid Compounds | | | | | |
| 2-Chlorophenol | 31 | 98 | 31 | 98 | 20 |
| 2,4 Dichlorophenol | 39 | 112 | 39 | 112 | 10 |
| 2,4 Dimethylphenol | 18 | 36 | 18 | 36 | - |
| 4,6 Dinitro-O-Cresol | NL | 60 | 78 | 277 | 60 |
| 2,4 Dinitrophenol | 71 | 123 | 71 | 123 | 40 |
| 2-Nitrophenol | 41 | 69 | 41 | 69 | - |
| 4-Nitrophenol | 72 | 124 | 72 | 124 | - |
| Pentachlorophenol | NL | 30 | NL | 30 | 30 |
| Phenol | 15 | 26 | 15 | 26 | 10 |
| 2,4,6 Trichlorophenol | NL | 20 | NL | 20 | 20 |
| Base/Neutral Compounds | | | | | |
| Anthracene | 22 | 59 | 22 | 59 | 10 |
| Benzidine | NL | 50 | NL | 50 | 50 |
| Benzo (a) Anthracene | NL | 10 | NL | 10 | 10 |
| Benzo (a) Pyrene | NL | 20 | NL | 20 | 20 |

| Parameter | FW2 Waters | | SE, SC Waters | | RQL* |
|-----------------------------------|------------|-----------|---------------|-----------|------|
| | Monthly / | Daily | Monthly / | Daily | |
| | Avg. | / Maximum | Avg. | / Maximum | |
| BaseNeutral Compounds (continued) | | | | | |
| Benzo (b) fluoranthene | NL | 10 | NL | 10 | 10 |
| Benzo (k) fluoranthene | NL | 20 | NL | 20 | 20 |
| Bis (2-Chloroethyl) Ether | NL | 10 | NL | 10 | 10 |
| Bis (2-Chloroisopropyl) Ether | 301 | 757 | 301 | 757 | 10 |
| Bis (2-Ethylhexyl)Phthalate | NL | 36 | 59 | 118 | 30 |
| Butyl Benzyl Phthalate | NL | 24 | NL | 24 | 20 |
| Chrysene | NL | 20 | NL | 20 | 20 |
| Dibenzo (a,h) Anthracene | NL | 20 | NL | 20 | 20 |
| 1,2 Dichlorobenzene | 77 | 163 | 77 | 163 | 9 |
| 1,3 Dichlorobenzene | 31 | 44 | 31 | 44 | 9 |
| 1,4 Dichlorobenzene | NL | 28 | NL | 28 | 20 |
| 3,3 Dichlorobenzidine | NL | 60 | NL | 60 | 60 |
| Diethyl Phthalate | 81 | 203 | 81 | 203 | 10 |
| Dimethyl Phthalate | 19 | 47 | 19 | 47 | 10 |
| Di-N-Butyl Phthalate | 27 | 57 | 27 | 57 | 20 |
| 2,4 Dinitrotoluene | NL | 10 | NL | 18.2 | 10 |
| 2,6 Dinitrotoluene | 255 | 641 | 255 | 641 | - |
| Fluoranthene | 25 | 68 | 25 | 68 | 10 |
| Fluorene | 22 | 59 | 22 | 59 | 10 |
| Hexachlorobenzene | NL | 10 | NL | 10 | 10 |
| Hexachlorobutadiene | NL | 10 | 20 | 49 | 10 |
| Hexchloropentadiene | 240 | 480 | NL | 1800 | 10 |
| Hexachloroethane | 19 | 38 | 21 | 54 | 10 |
| Ideno (1,2,3-cd) Pyrene | NL | 20 | NL | 20 | 20 |
| Isophorone | NL | 20 | NL | 20 | 10 |
| Nitrobenzene | 17 | 34 | 27 | 68 | 10 |
| N-Nitrosodimethylamine | NL | 20 | NL | 20 | 20 |
| N-Nitrosodiphenylamine | NL | 20 | NL | 20 | 20 |
| Phenanthrene | 22 | 59 | 22 | 59 | 10 |
| Pyrene | 25 | 67 | 25 | 67 | 20 |
| 1,2,4 Trichlorobenzene | 68 | 140 | 68 | 140 | 10 |
| Metals and Cyanide | | | | | |
| Arsenic ** | 50 | 100 | 50 | 100 | 8 |
| Cadmium ** | 50 | 100 | 50 | 100 | 4 |
| Chromium ** | 50 | 100 | 50 | 100 | 10 |
| Copper ** | 50 | 100 | 50 | 100 | 10 |
| Mercury ** | NL | 1 | NL | 1 | 1 |
| Nickel ** | 72 | 144 | 50 | 100 | 10 |
| Selenium ** | 50 | 100 | 50 | 100 | 10 |
| Silver ** | 25 | 50 | 25 | 50 | 2 |
| Zinc ** | 100 | 200 | 100 | 200 | 30 |
| Cyanide | 100 | 200 | 100 | 200 | 40 |

* The permittee shall ensure that analytical data is sampled at detection levels as sensitive as the Recommended Quantitation Levels (RQL's) for any of the above parameters limited in the individual authorization.

** If this parameter is regulated in the individual authorization, a chronic WET limit is also applicable.

Clarification of Reporting Requirements for MTBE Percent Removal

The equation for reporting MTBE percent removal is as follows:

$$\text{MTBE \% Removal} = \frac{\text{MTBE Influent} - \text{MTBE Effluent}}{\text{MTBE Influent}} \times 100$$

The equation above assumes that the permittee is monitoring MTBE influent and effluent only once during the calendar month as required in the master general permit. Permittees can always monitor more frequently than specified in the NJPDES permit as it encourages more representative data. In the event that a permittee monitors MTBE more than once during a particular calendar month, calculate individual percent removal values for each data set (influent and effluent) and divide by the number of data sets available to obtain the MTBE % removal value.

Note: Although examples are given below for all tables included in Part III of the master GPPC permit, discharges are typically authorized under only one table in individual authorizations. Therefore, only one of the five tables (A, B, C, D or E) of Part III is typically appropriate for a subject facility.

Given the above equation, reporting procedures are as follows for detectable and non-detectable or unquantified values:

- If the influent value is *detected and quantified* and the effluent value is *detected and quantified* and only one sample of each is taken during the calendar month, complete the calculation using the detected values. For example:

$$\text{MTBE \% Removal} = \frac{\text{MTBE Influent} - \text{MTBE Effluent}}{\text{MTBE Influent}} \times 100$$

MTBE Influent = 680 ug/L - only influent sample taken during the calendar month
MTBE Effluent = 90 ug/L - only effluent sample taken during the calendar month

$$\text{MTBE \% Removal} = \frac{680 - 90}{680} \times 100 = 86.7647 \quad \textbf{Report 87 \% on monitoring report form.}$$

- If the influent value is *detected and quantified* and the effluent value is *non-detectable or unquantified* and only one sample of each is taken during the calendar month, substitute one-half of the detection limit in the equation for the unquantified value. For example:

$$\text{MTBE \% Removal} = \frac{\text{MTBE Influent} - \text{MTBE Effluent}}{\text{MTBE Influent}} \times 100$$

MTBE Influent = 50 ug/L - only influent sample taken during the calendar month
MTBE Effluent = <5 ug/L - only effluent sample taken during the calendar month

$$\text{MTBE \% Removal} = \frac{50 - 2.5}{50} \times 100 = 95$$

For discharges authorized under Tables A, D and E: Because the effluent value is less than 70 ug/L, the MTBE minimum percent removal limitation does not apply, and **the permittee**

shall report “Code = N” on its monthly monitoring report form consistent with the instructions indicated in Part IV.

For discharges authorized under Tables B and C: Report 95% on monitoring report form.

- If the influent value is *non-detectable or unquantified* and the effluent value is *detected and quantified* and only one sample of each is taken during the calendar month, substitute one-half of the detection limit in the equation for the unquantified value. For example:

$$\text{MTBE \% Removal} = \frac{\text{MTBE Influent} - \text{MTBE Effluent}}{\text{MTBE Influent}} \times 100$$

MTBE Influent = <10 ug/L

- only influent sample taken during the calendar month

MTBE Effluent = 11 ug/L

- only effluent sample taken during the calendar month

$$\text{MTBE \% Removal} = \frac{5 - 11}{5} \times 100 = -120$$

For discharges authorized under Table A, D and E: Because the effluent value is less than 70 ug/L, the MTBE minimum percent removal limitation does not apply, and **the permittee shall report “Code = N”** on its monthly monitoring report form consistent with the instructions indicated in Part IV.

For discharges authorized under Tables B and C: Report -120%.

- If the influent value is *non-detectable or unquantified* and the effluent value is *non-detectable or unquantified* and only one sample of each is taken during the calendar month, substitute 0 for both influent and effluent values. For example:

$$\text{MTBE \% Removal} = \frac{\text{MTBE Influent} - \text{MTBE Effluent}}{\text{MTBE Influent}} \times 100$$

MTBE Influent = <10 ug/L

- only influent sample taken during the calendar month

MTBE Effluent = <5 ug/L

- only effluent sample taken during the calendar month

$$\text{MTBE \% Removal} = \frac{0 - 0}{0} \times 100 = 0$$

For discharges authorized under Table A, D and E: Because the effluent value is less than 70 ug/L, the MTBE minimum percent removal limitation does not apply, and **the permittee shall report “Code = N”** on its monthly monitoring report form consistent with the instructions indicated in Part IV.

For discharges authorized under Tables B and C: Report 0%.

APPENDIX A:

**CHRONIC TOXICITY TESTING SPECIFICATIONS
FOR USE IN THE NJPDES PERMIT PROGRAM**

Version 2.1

May 1997

(This Section is only applicable if a chronic toxicity limit is required as specified in Part III. Chronic toxicity limits are applied only under Table D and if metals are present at certain levels.)

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VIII. REFERENCES

Notice: Mention of trade names or commercial products do not constitute endorsement or recommendation for use.

I. AUTHORITY AND PURPOSE

These methods specifications for the conduct of whole effluent chronic toxicity testing are established under the authority of the NJPDES permitting program, N.J.A.C. 7:14A-6.5(a)2 and 40 CFR 136, for discharges to waters of the State. The methods referenced herein are included by reference in 40 CFR 136, Table 1.A. and, therefore, constitute approved methods for chronic toxicity testing. The information contained herein serves to clarify testing requirements not sufficiently clarified in those methods documents and also serves to outline and implement the interlaboratory Standard Reference Toxicant Program until a formal laboratory certification program is established under N.J.A.C. 7:18. As such these methods are intended to be used to determine compliance with discharge permits issued under the authority of the NJPDES permit program. Tests are to be conducted in accordance with the general conditions and test organism specific method specifications contained in this document. All other conditions and specifications can be found in 40 CFR 136 and USEPA methodologies.

Until a subchapter on chronic toxicity testing within the regulations governing the certification of laboratories and environmental measurements (N.J.A.C. 7:18) becomes effective, tests shall be conducted in conformance with the methodologies as designated herein and contained in 40 CFR 136. The laboratory performing the testing shall be within the existing acute toxicity testing laboratory certification program established under N.J.A.C. 7:18, as required by N.J.A.C. 7:9B-1.5(c)5.

Testing shall be in conformance with the subchapter on chronic toxicity testing within the N.J.A.C. 7:18 when such regulations become effective. The laboratory performing the toxicity testing shall be within the chronic toxicity testing laboratory certification program to be established under that subchapter, when it becomes effective.

These methods are incorporated into discharge permits as enforceable permit conditions. Each discharge permit will specify in Part IV of the permit, the test species specific methods from this document that will be required under the terms of the discharge permit. Although the test species specific methods for each permit are determined on a case-by-case basis, the purpose of this methods document is to assure consistency among dischargers and to provide certified laboratories with information on the universe of tests to be utilized so that they can make the necessary preparations, including completing the required Standard Reference Toxicant testing. Please note that these methodologies are required for compliance testing only. Facilities and/or laboratories conducting testing under the requirements of a Toxicity Identification Evaluation or for informational purposes are not bound by these methods.

This document constitutes the second version of the NJDEP's interim chronic methodologies. This version contains no significant changes to the test methods themselves. However, in keeping with the Department's continued emphasis on good laboratory practices and quality control, the areas addressing the Standard Reference Toxicant Program, data analysis and data reporting, have been significantly revised.

II. GENERAL CONDITIONS

A. LABORATORY SAFETY, GLASSWARE, ETC.

All safety procedures, glassware cleaning procedures, etc., shall be in conformance with 40 CFR 136 and USEPA's "Short Term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Freshwater Organisms," "Short Term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Marine and Estuarine Organisms" and N.J.A.C. 7:18.

B. TEST CONCENTRATIONS / REPLICATES

All testing is to be performed with a minimum of five effluent concentrations plus a dilution water control. A second reference water control is optional when a dilution water other than culture water is used. The use of both a 0.5 or 0.75 dilution factor is acceptable for the selection of test concentrations. If hypothesis testing will be used to determine the test endpoint, one effluent concentration shall be the chronic permit limitation, unless the existing data for the discharge indicate that the NOEC is expected to be significantly less than the permit limit. The use of the 0.5 dilution factor may require more than five dilutions to cover the entire range of effluent concentrations as well as the chronic permit limit, since the permit limit will often not be one of the nominal concentrations in a 0.5 dilution series. In such an instance, the 0.5 dilution series may be altered by including an additional test concentration equal to the permit limit in the dilution series, or by changing the concentration closest to the permit toxicity limit to be equal to that limit. The Department recommends the use of the 0.75 dilution factor using Table 1.0 to determine test concentrations. That table establishes test concentrations based on the chronic toxicity limitation.

For either the 0.5 or 0.75 dilution factor, there shall be at least one test concentration above the permit limitation and at least three test concentrations below the permit limit along with the dilution water control unless the permit limitation prohibits such (e.g., limitations greater than 75% effluent). An effort shall be made to bracket the anticipated test result.

To use Table 1.0, locate the permit limit in column 4. The dilution series becomes the row that corresponds to the permit limit in column 4. For example, a permit limit of 41 would require a dilution series of the dilution water control, 17%, 23%, 31%, 41% and 55% effluent.

The number of replicates used in the test must, at a minimum, satisfy the specifications of the applicable methods contained herein. Increased data sensitivity can be obtained by increasing the number of replicates equally among test concentrations and thus an increased number of replicates is acceptable. Further, the use of nonparametric statistical analysis requires a minimum of four replicates per test concentration. If the data for any particular test is not conducive to parametric analyses and if less than four replicates were included, the test may not be considered acceptable for compliance purposes.

The use of single concentration tests consisting of the permit limitation as a concentration and a control is not permitted for compliance purposes, but may be used by a permittee in the conduct of a Toxicity Investigation Evaluation (TIE) or for information gathering purposes. Such a test would be considered a "pass" if there was no significant difference in test results, using hypothesis testing methods.

Table 1.0: 0.75 DILUTION SERIES INDEXED BY PERMIT LIMIT

| | | | | Permit Limit | | | | | | Permit Limit | |
|-------|-----|-----|-----|--------------|-----|-------|----|----|----|--------------|-----|
| Col # | 1 | 2 | 3 | 4 | 5 | Col # | 1 | 2 | 3 | 4 | 5 |
| | 0.4 | 0.6 | 0.8 | 1 | 1.3 | | 22 | 29 | 38 | 51 | 68 |
| | 0.8 | 1.1 | 1.5 | 2 | 2.7 | | 22 | 29 | 39 | 52 | 69 |
| | 1.3 | 1.7 | 2.3 | 3 | 4 | | 22 | 30 | 40 | 53 | 71 |
| | 1.7 | 2.3 | 3 | 4 | 5.3 | | 23 | 30 | 41 | 54 | 72 |
| | 2.1 | 2.8 | 3.8 | 5 | 6.7 | | 23 | 31 | 41 | 55 | 73 |
| | 2.5 | 3.4 | 4.5 | 6 | 8 | | 24 | 32 | 42 | 56 | 75 |
| | 3 | 4 | 5 | 7 | 9 | | 24 | 32 | 43 | 57 | 76 |
| | 3 | 5 | 6 | 8 | 11 | | 24 | 33 | 44 | 58 | 77 |
| | 4 | 5 | 7 | 9 | 12 | | 25 | 33 | 44 | 59 | 79 |
| | 4 | 6 | 8 | 10 | 13 | | 25 | 34 | 45 | 60 | 80 |
| | 5 | 6 | 8 | 11 | 15 | | 26 | 34 | 46 | 61 | 81 |
| | 5 | 7 | 9 | 12 | 16 | | 26 | 35 | 47 | 62 | 83 |
| | 5 | 7 | 10 | 13 | 17 | | 27 | 35 | 47 | 63 | 84 |
| | 6 | 8 | 11 | 14 | 19 | | 27 | 36 | 48 | 64 | 85 |
| | 6 | 8 | 11 | 15 | 20 | | 27 | 37 | 49 | 65 | 87 |
| | 7 | 9 | 12 | 16 | 21 | | 28 | 37 | 50 | 66 | 88 |
| | 7 | 10 | 13 | 17 | 23 | | 28 | 38 | 50 | 67 | 89 |
| | 8 | 10 | 14 | 18 | 24 | | 29 | 38 | 51 | 68 | 91 |
| | 8 | 11 | 14 | 19 | 25 | | 29 | 39 | 52 | 69 | 92 |
| | 8 | 11 | 15 | 20 | 27 | | 30 | 39 | 53 | 70 | 93 |
| | 9 | 12 | 16 | 21 | 28 | | 30 | 40 | 53 | 71 | 95 |
| | 9 | 12 | 17 | 22 | 29 | | 30 | 41 | 54 | 72 | 96 |
| | 10 | 13 | 17 | 23 | 31 | | 31 | 41 | 55 | 73 | 97 |
| | 10 | 14 | 18 | 24 | 32 | | 31 | 42 | 56 | 74 | 99 |
| | 11 | 14 | 19 | 25 | 33 | | 32 | 42 | 56 | 75 | 100 |
| | 11 | 15 | 20 | 26 | 35 | 24 | 32 | 43 | 57 | 76 | |
| | 11 | 15 | 20 | 27 | 36 | 24 | 32 | 43 | 58 | 77 | |
| | 12 | 16 | 21 | 28 | 37 | 25 | 33 | 44 | 59 | 78 | |
| | 12 | 16 | 22 | 29 | 39 | 25 | 33 | 44 | 59 | 79 | |
| | 13 | 17 | 23 | 30 | 40 | 25 | 34 | 45 | 60 | 80 | |
| | 13 | 17 | 23 | 31 | 41 | 26 | 34 | 46 | 61 | 81 | |
| | 14 | 18 | 24 | 32 | 43 | 26 | 35 | 46 | 62 | 82 | |
| | 14 | 19 | 25 | 33 | 44 | 26 | 35 | 47 | 62 | 83 | |
| | 14 | 19 | 26 | 34 | 45 | 27 | 35 | 47 | 63 | 84 | |
| | 15 | 20 | 26 | 35 | 47 | 27 | 36 | 48 | 64 | 85 | |
| | 15 | 20 | 27 | 36 | 48 | 27 | 36 | 48 | 65 | 86 | |
| | 16 | 21 | 28 | 37 | 49 | 28 | 37 | 49 | 65 | 87 | |
| | 16 | 21 | 29 | 38 | 51 | 28 | 37 | 50 | 66 | 88 | |
| | 16 | 22 | 29 | 39 | 52 | 28 | 38 | 50 | 67 | 89 | |
| | 17 | 23 | 30 | 40 | 53 | 28 | 38 | 51 | 68 | 90 | |
| | 17 | 23 | 31 | 41 | 55 | 29 | 38 | 51 | 68 | 91 | |
| | 18 | 24 | 32 | 42 | 56 | 29 | 39 | 52 | 69 | 92 | |
| | 18 | 24 | 32 | 43 | 57 | 29 | 39 | 52 | 70 | 93 | |
| | 19 | 25 | 33 | 44 | 59 | 30 | 40 | 53 | 71 | 94 | |
| | 19 | 25 | 34 | 45 | 60 | 30 | 40 | 53 | 71 | 95 | |
| | 19 | 26 | 35 | 46 | 61 | 30 | 41 | 54 | 72 | 96 | |
| | 20 | 26 | 35 | 47 | 63 | 31 | 41 | 55 | 73 | 97 | |
| | 20 | 27 | 36 | 48 | 64 | 31 | 41 | 55 | 74 | 98 | |
| | 21 | 28 | 37 | 49 | 65 | 31 | 42 | 56 | 74 | 99 | |
| | 21 | 28 | 38 | 50 | 67 | 32 | 42 | 56 | 75 | 100 | |

* Select the dilution series by finding the row which contains the permit limit in column #4.

NOTE: All values are in units of "% effluent" not toxic units.

C. DILUTION WATER

1. Marine and Estuarine Waters

A high quality natural water, such as the Manasquan River Inlet is strongly recommended as the dilution water source for chronic toxicity testing with marine and estuarine organisms. The use of the receiving water as the dilution water source is not required. Saline waters prepared with hypersaline brine and deionized water may also be used as dilution water. Hypersaline brines shall be prepared from a high quality natural seawater and shall not exceed a concentration of 100 ppt. The type of a dilution water for a permittee may not be changed without the prior approval of the Department.

The standard test salinity shall be 25 ppt, except for *Champia parvula*, which shall be tested at 30 ppt. Since most effluents are freshwater based, in most cases it will be necessary to adjust the salinity of the test concentrations to the standard test salinity.

2. Fresh Waters

A high quality natural water, such as Round Valley Reservoir (if access is allowed) or Lake Hopatcong, is strongly recommended as the dilution water source for chronic toxicity testing with freshwater organisms. It is not required to perform the toxicity testing with the receiving water as dilution water. Tests performed with a reconstituted water or up to 20% Diluted Mineral Water (DMW) as dilution water is acceptable. For testing with *Ceriodaphnia dubia*, the addition of 5 µg/l selenium (2 µg/l selenium with natural water) and 1 µg/l vitamin B12 is recommended (Keating and Dagbusan, 1984; Keating, 1985 and 1988). The source of a dilution water for a permittee may not be changed without the prior approval of the Department. Reconstituted water and DMW should be prepared with Millipore Super Q^R or equivalent, meet the requirements of N.J.A.C. 7:18-6 and should be aerated a minimum of 24 hrs prior to use, but not supersaturated.

D. EFFLUENT SAMPLE COLLECTION

Effluent samples shall be representative of the discharge being regulated. For each discharge serial number (DSN), the effluent sampling location shall be the same as that specified in the NJPDES permit for other sampling parameters unless an alternate sampling point is specified in the NJPDES discharge permit. For industrial dischargers with a combined process/sanitary waste stream, effluent sampling shall be after chlorination, unless otherwise designated in the permit.

For continuous discharges, effluent sampling shall consist of 24 hour composite samples consisting either of equal volumes taken once every hour or of a flow-proportionate composite sample, unless otherwise approved by the Department. At a minimum, three samples shall be collected as specified above, one every other day. The first sample shall be used for test initiation and the first renewal. The second sample for the next two renewals. The third sample shall be used for the final three renewals. For the *Champia* and *Selenastrum* tests, a single sample shall be collected not more than 24 hours prior to test initiation. No effluent sample shall be over 72 hours old at the time of its use to initiate or renew solutions in a test. It is acceptable to collect samples more frequently for chronic WET testing and if samples are collected daily for acute toxicity testing conducted concurrently, available samples may be used to renew the test solutions as appropriate.

For all other types of discharges, effluent sampling shall be conducted according to specifications contained within the discharge permit, methodology questionnaire or as otherwise specified by the Department. The use of grab samples or other special sampling procedures will be based on time of occurrence and duration of intermittent discharge events.

If a municipal discharger has concerns that the concentrations of ammonia and/or chlorine in an effluent are adequate to cause violations of the permit limit for chronic toxicity testing, the permittee should conduct analyses, as specified in USEPA's toxicity investigation methods documents, to illustrate the relationship between chronic effluent toxicity and chlorine and/or ammonia as applicable. This data may then be submitted to the Department

as justification for a request to use modified test procedures, which account for ammonia and/or chlorine toxicity, in future chronic toxicity tests. The Department may, where adequate justification exists, permit the adjustment of these pollutants in the effluent sample if discharge limits for these pollutants are contained in the NJPDES permit and those permit limitations are adequate for the protection of water quality. Any proposed modified test procedures to adjust effluent chlorine and/or ammonia shall be approved by the Department prior to use of those test procedures for any compliance testing.

Except for filtration through a 2 mm or larger screen or an adjustment to the standard test salinity, no other adjustments to the effluent sample shall be made without prior written approval by the Department. Aeration of samples prior to test start shall be minimized where possible and samples shall not be aerated where adequate saturation exists to maintain dissolved oxygen.

E. PHYSICAL CHEMICAL MEASUREMENTS

At a minimum, the physical chemical measurements shall be as follows:

- pH and dissolved oxygen shall be measured at the beginning and end of each 24 hour exposure period, in at least one chamber, of the high, medium and low test concentrations and the control. In order to ensure that measurements for these parameters are representative of the test concentrations during the test, measurements for these parameters should be taken in an additional replicate chamber for such concentrations which contains no test organisms, but is subject to the same test conditions.
- Temperature shall either be monitored continuously, measured daily in at least two locations in the environmental control system, or measured at the beginning of each 24 hr exposure period in at least one replicate for each treatment.
- Salinity shall be measured in all salt water tests at the beginning of each 24 hour exposure period, in at least one replicate for each treatment.
- For all freshwater tests, alkalinity, hardness and conductivity shall be measured in each new sample (100% effluent) and control.
- Nitrite, nitrate and ammonia shall be measured in the control before each renewal in the mysid test only.
- For samples of discharges where concentrations of ammonia and/or chlorine are known or are suspected to be sufficient to cause toxicity, it is recommended that the concentrations of these pollutants be determined and submitted with the standardized report form. The laboratory is advised to consult with the permittee to determine if these parameters should be measured in the effluent. Where such measurements are deemed appropriate, measurements shall be conducted at the beginning of each 24 hour exposure period. Also, since a rise in the test pH can affect the toxicity of ammonia in the effluent, analysis of ammonia during the test may be appropriate if a rise in pH is accompanied by a significant increase in mortality.

F. STATISTICS

The use of both hypothesis testing techniques and point estimate techniques are currently in use by the Department or by permittees for compliance purposes. The NJPDES permit should be checked to determine which type of analysis is required and appropriate for each specific facility. It is not acceptable to simply evaluate any data by "visual data review" unless in the analysis of survival data, no mortality occurred in the test. All data sets must be appropriately statistically evaluated.

For hypothesis testing techniques, statistical analysis shall follow the protocols in USEPA (1988, 1989) to evaluate adverse effects. A significance level of 0.05 shall be utilized to evaluate such effects. Use of a protocol

not contained in these documents must be accompanied by a reference and explanation addressing its applicability to the particular data set. Please note the following when evaluating data using hypothesis testing techniques.

Special attention should be given to the omission and inclusion of a given replicate in the analysis of mysid fecundity data (USEPA 1994, p. 275) and *Ceriodaphnia* reproduction data (USEPA 1994, page 174).

Determination of acceptability criteria and average individual dry weight for the growth endpoints must follow the specifications in the applicable documents (e.g., p.84 for saltwater methods document.)

Use of nonparametric statistical analyses requires a minimum of four replicates per test concentration. If the data for any particular test are not conducive to parametric analyses and if less than four replicates were included, the test may not be acceptable to the Department.

Where hypothesis testing is used for compliance purposes, if the results of hypothesis testing indicate that a deviation from the dose response occurs such that two test concentrations are deemed statistically significant from the control but an intermediate test concentration is not, the test is deemed unacceptable and cannot be used for compliance testing purposes.

For point estimate techniques, statistical analysis should follow the protocol contained in "A Linear Interpolation Method for Sublethal Toxicity: The Inhibition Concentration (ICp) Approach (Version 2.0), July 1993, National Effluent Toxicity Assessment Center Technical Report 03-93." Copies of the program can be obtained by contacting the Department. The linear interpolation estimate ICp values and not the bootstrap mean ICp, shall be reported for permit compliance purposes. The ICp value reported on the Discharge Monitoring Report shall be rounded off as specified in the Department's "Discharge Monitoring Report (DMR) Instruction Manual, December 1993." IC25 values shall be reported under the parameter code listed as "NOEC" on the DMR, until the DMR's are adjusted accordingly.

If the result reported by the ICp method is greater than the highest concentration tested, the test result is reported as "greater than C" where "C" is the highest tested concentration. If the ICp is lower than the lowest concentration tested, the test result is reported as "less than C" where "C" is the lowest tested concentration.

If separate NOEC's/IC25's can be calculated from multiple test endpoints, for example a reproductive endpoint and a growth endpoint, the lowest NOEC/IC25 value expressed in units of "% effluent" will be used to determine permit compliance and should, therefore, be reported as the NOEC/IC25 value for the test. If the NOEC value for growth and/or reproduction is not lower than that for survival, the NOEC/IC25 value reported for the test shall be as survival. For saltwater tests, where additional controls are used in a test (i.e. brine and/or artificial sea salt control), a T-test shall be used to determine if there is a significant difference between the original test control and the additional controls. If there is a significant difference between any of the controls, the test may be deemed unacceptable and if so, will not be used for permit compliance.

III. TEST ACCEPTABILITY CRITERIA

Any test that does not meet these acceptability criteria will not be used by the Department for any purpose and must be repeated as soon as practicable, with a freshly collected sample.

1. Tests must be performed by a laboratory approved for the conduct of chronic toxicity tests and certified for acute toxicity testing under N.J.A.C. 7:18.
2. Test results may be rejected due to inappropriate sampling, including the use of less than three effluent samples in a test and/or use of procedures not specified in a permit or methodology questionnaire, use of frozen or unrefrigerated samples or unapproved pretreatment of an effluent sample.
3. Controls shall meet the applicable performance criteria specified in the Table 2.0 and in the individual method specifications contained herein.
4. Acceptable and applicable Standard Reference Toxicant Data must be available for the test.
5. No unapproved deviations from the applicable test methodology may be present.
6. When using hypothesis testing techniques, a deviation from the dose response as explained in the statistical portion of this document shall not be present in the data.

Table 2.0: CONTROL PERFORMANCE

| TEST ORGANISM | MINIMUM SURVIVAL | MINIMUM WEIGHT GAIN | MINIMUM FECUNDITY/ REPRODUCTION |
|----------------------------------|---------------------------------------|------------------------------------------------------|-------------------------------------------------------------------------------------------------------------------------------------------------------|
| <i>Pimephales promelas</i> | 80% | 0.25 mg avg | N/A |
| <i>Ceriodaphnia dubia</i> | 80% | N/A | Average of ≥ 15 young per surviving female |
| <i>Selenastrum capricornutum</i> | Density $\geq 2 \times 10^5$ cells/ml | N/A | Variability in controls not to exceed 20%. |
| <i>Cyprinodon variegatus</i> | 80% | 0.60 mg (unpreserved) avg 0.50 mg (preserved) avg | N/A |
| <i>Menidia beryllina</i> | 80% | 0.50 mg (unpreserved) avg 0.43 mg (preserved) avg | N/A |
| <i>Mysidopsis bahia</i> | 80% | 0.2 mg per mysid avg | egg production by 50% of control females if fecundity is used as an endpoint. |
| <i>Champia parvula</i> | 100% | N/A | ≥ 10 cystocarps per plant Plants in controls and lower test concentrations shall not fragment so that individual plants cannot be identified. |

THE DETERMINATION OF A TEST AS UNACCEPTABLE DOES NOT RELIEVE THE FACILITY FROM MONITORING FOR THAT MONITORING PERIOD

IV. STANDARD REFERENCE TOXICANT TESTING

All chronic testing shall be accompanied by testing with a Standard Reference Toxicant (SRT) as a part of each laboratory's internal quality control program. Such a testing program should be consistent with the quality assurance/quality control protocols described in the USEPA chronic testing manuals. Laboratories may utilize the reference toxicant of their choice and toxicants such as cadmium chloride, potassium chloride, sodium dodecyl sulfate and copper sulfate are all acceptable. However, Potassium chloride has been chosen by several laboratories and is recommended by the Department. The concentration of the reference toxicant shall be verified by chemical analysis in the low and high test concentrations once each year or every 12 tests, whichever is less. It is not necessary to run SRT tests, for all species using the same SRT.

A. INITIAL STANDARD REFERENCE TOXICANT (SRT) TESTING REQUIREMENTS

At a minimum, this testing shall include an initial series of at least five SRT tests for each test species method. Acceptable SRT testing for chronic toxicity shall be performed utilizing the short term chronic toxicity test methods as specified herein. Reference toxicant tests utilizing acute toxicity testing methods, or any method other than those contained in this document are not acceptable. The laboratory should forward results of the initial SRT testing, including control charts, the name of the reference toxicant utilized, the supplier and appropriate chemical analysis of the toxicant to either address listed in the reporting requirements section herein.

The initial series of a least five SRT tests for a specific test species method shall be completed and approved in writing by the Department prior to the conduct of any chronic toxicity testing for compliance purposes.

B. SUBSEQUENT SRT TESTING REQUIREMENTS

After receiving the initial approval from the Department to conduct chronic toxicity tests for compliance purposes, subsequent SRT testing shall be conducted as follows:

1. Where organisms used in testing are cultured at the testing laboratory, SRT testing should be conducted once per month for each species/method.
2. Where the laboratory purchases organisms from a laboratory certified in New Jersey for the conduct of acute toxicity testing and approved for the conduct of chronic toxicity testing for the test organism in question (i.e. the "supplier laboratory"), SRT data provided by the "supplier laboratory" for each lot of organisms purchased is acceptable as long as the SRT test result falls within the control limits of the control chart established by the "supplier laboratory" for that organism. The laboratory using purchased organisms is responsible for the results of any compliance tests they perform.
3. A testing laboratory purchasing organisms from a supplier laboratory must still perform SRT testing on a quarterly basis at a minimum, for each species they test with, in order to adequately document their own interlaboratory precision.
4. If a testing laboratory purchasing organisms elects not to use the SRT data from a "supplier laboratory" or such data is unavailable or where organisms are purchased from another organism supplier, the testing laboratory must conduct SRT testing on each lot of organisms purchased.
5. For industrial laboratories certified under N.J.A.C. 7:18 to conduct acute toxicity tests, only the SRT testing conditions specified in 2. through 4. above apply. Where that laboratory/facility cultures their own test organisms, the frequency of SRT testing required will be determined on a case by case basis, based on the frequency of testing for that facility.

NOTE: Based on these requirements, SRT data are considered applicable to a compliance test when the SRT test results are acceptable and the SRT test is conducted within 30 days of the compliance test, for the test species and SRT in question. Therefore, it is not necessary for an approved laboratory to run an SRT test every month if the laboratory is not conducting compliance tests for a particular species.

C. CHANGING OF AN ESTABLISHED REFERENCE TOXICANT

The SRT used for any species by a laboratory may be changed at any time provided that the following conditions have been satisfied:

1. A series of at least three reference toxicant tests are conducted with the new reference toxicant and the results of those tests are identified as satisfactory, in writing, by the Department.
2. Laboratories must continue using the already approved SRT in their ongoing QA/QC program, until such time as the letter referenced above, is received by the laboratory.

D. CONTROL CHARTS

Control charts shall be established from SRT test results in accordance with the procedures outlined in the USEPA methods documents. Control charts shall be constructed using IC25's using the following methods:

1. The upper and lower control limits shall be calculated by determining +/- two standard deviations above and below the mean.
2. SRT test results which exhibit an IC25 that is greater than the highest concentration tested or less than the lowest concentration tested (i.e. a definitive endpoint cannot be determined), shall not be used to establish control charts.
3. SRT tests which do not meet the acceptability criteria for a specific species shall not be used to establish control charts.
4. All values used in the control charts should be as nominal concentrations. However, the control charts shall be accompanied by a chart tabulating the test results as measured concentrations.
5. An outlier (i.e. values which fall outside the upper and lower control limits) should be included on the control chart unless it is determined that the outlier was caused by factors not directly related to the test organisms (e.g., test concentration preparation) as the source of variability would not be directly applicable to effluent tests. In such case, the result and explanation shall be reported to the Department within 30 days of the completion of the SRT test.

The control chart established for the initial series of SRT data submitted will be used by the laboratory and the Department to determine outliers from SRT test results reported in the "NJPDES Biomonitoring Report Form - Chronic Toxicity Test" submitted by the permittees for the test species. These initial control limits will remain unchanged until twenty SRT tests have been completed by the laboratory.

The following procedures shall be used for continually updating control charts after twenty acceptable SRT tests have been completed:

1. Once a laboratory has completed twenty acceptable SRT tests for a test species, the upper and lower control limits shall be recalculated with those twenty values.
2. For each successive SRT test conducted after these first twenty tests, a moving average shall be calculated and the control limits reevaluated using the last twenty consecutive test results.
3. The upper and lower control limits shall be reported on the "NJPDES Biomonitoring Report Form - Chronic Toxicity Tests" along with the SRT test result.

E. UNACCEPTABLE SRT TEST RESULTS

If a laboratory produces any SRT test results which are outside the established upper and lower control limits for a test species at a frequency greater than one test in any ten tests, a report shall be forwarded to the Department at the address contained herein. This report shall include any identified problem which caused the values to fall outside the expected range and the corresponding actions that have been taken by the laboratory. The Department may not accept or may require repeat testing for any toxicity testing that may have been affected by such an occurrence.

If a laboratory produces two consecutive SRT test results or three out of any ten test results which are outside the established upper and lower limits for a specific test species, the laboratory shall be unapproved to conduct chronic toxicity tests for compliance purposes for that test species. Reapproval shall be contingent upon the laboratory producing SRT test results within the established upper and lower control limits for that test species in two consecutive SRT tests. If one or both of those test results again fall outside the established control levels, the laboratory is unapproved for that test species until five consecutive test results within the established upper and lower control limits are submitted and approved by the Department.

F. ANNUAL SUBMITTALS

Control charts shall be forwarded to the Department on an annual basis, on the anniversary of approval for the test species.

The Department may request, at any time, any information which is essential in the evaluation of SRT results and/or compliance data.

V. TEST CANCELLATION / RESCHEDULING EVENTS

A lab may become aware of QA problems during or immediately following a test that will prevent data from being submitted or a lab may be unable to complete a tests due to sample collection or shipping problems. If for any reason a chronic toxicity test is initiated and then prematurely ended by the laboratory or at the request of the permittee, the laboratory shall submit the form entitled "Chronic Whole Effluent Toxicity Testing Test Cancellation / Rescheduling Event Form" contained herein. This form shall be used to detail the reason for prematurely ending the test. This completed form and any applicable raw data sheets shall be submitted to the appropriate biomonitoring program at the address above within 30 days of the cessation of the test.

Tests are considered to be initiated once test organisms have been added to all test chambers.

Submission of this form does not relieve the facility from monitoring for that monitoring period.

VI. REPORTING

The report form entitled "NJPDES Biomonitoring Report Form - Chronic Toxicity Tests" should be used to report the results of all NJPDES chronic compliance biomonitoring tests. Laboratory facsimiles are acceptable but must contain all information included on any recent revisions of the form by the Department. Statistical printouts and raw data sheets for all endpoints analyzed shall be included with the report submitted to the Department. Two copies of all chronic toxicity test report forms shall be submitted to the following address as applicable:

Bureau of Point Source Permitting Region 1 **OR**
Bureau of Point Source Permitting Region 2 (as indicated in the cover letter)

New Jersey Department of Environmental Protection
Division of Water Quality
PO Box 29
Trenton, NJ 08625-0029

It is not necessary to attach a copy of a test report form to the Discharge Monitoring Report (DMR) form when submitting this form to the Department. However, the results of all chronic toxicity tests conducted for compliance purposes must be reported on the DMR form under the appropriate parameter code in the monitoring period in which the test was conducted.

VII. METHOD SPECIFICATIONS

The following method specifications shall be followed as specified in the NJPDES permit. Any changes to these methods will not be considered acceptable unless they are approved in writing by the Department, prior to their use.

- A. Fathead Minnow (*Pimephales promelas*), Larval Survival and Growth Test, method 1000.0
- B. *Ceriodaphnia dubia*, Survival and Reproduction Test, method 1002.0
- C. Algal, (*Selenastrum capricornutum*), Growth Test, method 1003.0
- D. Sheepshead Minnow (*Cyprinodon variegatus*), Larval Survival and Growth Test, method 1005.0
- E. Inland Silverside (*Menidia beryllina*), Larval Survival and Growth Test, method 1006.0
- F. *Mysidopsis bahia*, Survival, Growth, and Fecundity Test, method 1007.0
- G. *Champia parvula*, Sexual Reproduction Test, method 1009.0

VIII. REFERENCES

1. Keating, K. 1985. The influence of Vitamin B12 deficiency on the reproduction of Daphnia pulex Leydig (Cladocera). J. Crustacean Biology 5:130-136.
2. Keating, K. 1988. N.J.D.E.P. Project C29589, Fiscal 1988 Third Quarter Summary Report. Producing Nutritionally Competent Daphnids for Use in Bioassay. 44p.
3. Keating, K., and B. Dagbusan. 1984. Effect of selenium deficiency on cuticle integrity in Cladocera (Crustacea). Proc. Natl. Acad. Sci. USA 81:3433-3437.
4. NJDEP, 1993. Discharge Monitoring Report (DMR) Instruction Manual.
5. USEPA. 1994. Short Term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Marine and Estuarine Organisms. EPA-600/4-91-003. July 1994. Second Edition.
6. USEPA. 1994. Short Term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Freshwater Organisms. EPA/600/4-91/002. July 1994. Third Edition.

EFFLUENT SAMPLING:

SAMPLING POINT / DESCRIPTION OF SAMPLING SITE:

SAMPLING INITIATED: DATE: ____/____/____ TIME: _____

SAMPLING ENDED: DATE: ____/____/____ TIME: _____

NUMBER OF EFFLUENT SAMPLES COLLECTED: _____

SAMPLE TYPE (GRAB/COMPOSITE): _____

RECEIVED IN LAB BY/FROM:

METHOD OF SHIPMENT:

(ALL APPLICABLE RAW DATA SHEETS MUST BE ATTACHED)
c: Permittees authorized agent.